



Genome Report

Complete Whole-Genome Sequence of *Streptomyces* sp. MUM 178J, a Potential Anti-*Vibrio* Agent

Ke-Yan Loo^{1,2}, Loh Teng-Hern Tan^{1,3*}, Kah-Ooi Chua⁴, Priyia Pusparajah⁵, Kok-Gan Chan⁶, Learn-Han Lee^{1,7*}, Jodi Woan-Fei Law^{1,8*}, Vengadesh Letchumanan^{1,2}

Article History Received: 21 December	¹ Microbiome and Bioresource Research Strength (MBRS), Jeffrey Cheah School of Medicine and Health Sciences, Monash University Malaysia, Bandar Sunway 47500, Selangor Darul Ehsan, Malaysia; ke.loo@monash.edu (K-YL)
2023; Received in Revised Form: 03 February 2024;	² Pathogen Resistome Virulome and Diagnostic Research Group (PathRiD), Jeffrey Cheah School of Medicine and Health Sciences, Monash University Malaysia, Bandar Sunway 47500, Selangor Darul Ehsan, Malaysia; vengadesh.letchumanan1@monash.edu (VL)
Accepted: 14 February 2024; Available Online: 16 February 2024	³ Innovative Bioprospection Development Research Group (InBioD), Research Center for Life Science and Healthcare, China Beacons of Excellence Research and Innovation Institute (CBI), University of Nottingham Ningbo China, Zhejiang, China
	⁴ Centre for Research in Biotechnology for Agriculture (CEBAR), University of Malaya, Kuala Lumpur 50603, Malaysia; kahooi@um.edu.my (K-OC)
	⁵ Medical Health and Translational Research Group (MHTR), Faculty of Medicine, Nursing and Health Sciences, Monash University, Clayton, Melbourne, Australia; priyia.pusparajah@monash.edu (PP)
	⁶ Division of Genetics and Molecular Biology, Institute of Biological Sciences, Faculty of Science, University of Malaya, Kuala Lumpur 50603, Malaysia; kokgan@um.edu.my (K-GC)
	⁷ Novel Bacteria and Drug Discovery Research Group (NBDD), Research Center for Life Science and Healthcare, China Beacons of Excellence Research and Innovation Institute (CBI), University of Nottingham Ningbo China, Zhejiang, China
	⁸ Next-Generation Precision Medicine and Therapeutics Research Group (NMeT), Research Center for Life Science and Healthcare, China Beacons of Excellence Research and Innovation Institute (CBI), University of Nottingham Ningbo China, Zhejiang, China
	*Corresponding author: Loh Teng Hern-Tan; Innovative Bioprospection Development Research Group (InBioD), Research Center for Life Science and Healthcare, China Beacons of Excellence Research and Innovation Institute (CBI), University of Nottingham Ningbo China, Zhejiang, China; loh.teng.hern@monash.edu (LT-HT); Learn-Han Lee; Novel Bacteria and Drug Discovery Research Group (NBDD), Research Center for Life Science and Healthcare, China Beacons of Excellence Research and Innovation Institute (CBI), University of Nottingham Ningbo China, Zhejiang, China; Learn-Han.Lee@nottingham.edu.cn (L-HL); Jodi Woan-Fei Law; Next- Generation Precision Medicine and Therapeutics Research Group (NMeT), Research Center for Life Science and Healthcare, China Beacons of Excellence Research and Innovation Institute (CBI), University of Nottingham Ningbo China, Zhejiang, China; jodi.law1@monash.edu (JW-FL)

Abstract: Streptomyces sp. is a group of filamentous, Gram-positive bacteria well known for their capabilities in producing bioactive compounds that have been used as novel drugs and lead in drug development. The Streptomyces sp. MUM 178J was isolated from a mangrove forest in Malaysia. This isolate was found to harbor anti-Vibrio properties as its crude extract inhibited the growth of multidrug-resistant Vibrio parahaemolyticus. Therefore, the strain was subjected to whole genome sequencing to unearth its genomic potential. The genome of Streptomyces sp. MUM 178J consists of 6,699,249 bp with a G+C content of 71.3%. 66 tRNA genes and 18 rRNA genes were also predicted to be present within the genome. Further analysis with the bioinformatics tool, antiSMASH (antibiotics & Secondary Metabolite Analysis Shell), detected nine biosynthetic gene clusters displaying more than 70% similarity to known gene clusters, including one associated with melanin production. Melanin has demonstrated antagonistic activity against the growth of members of the Vibrio family, including V. parahaemolyticus. This indicates the potential correlation between the production of melanin and the anti-Vibrio properties of MUM 178J. The availability of the whole genome sequence of Streptomyces sp. MUM 178J allows for future in-depth investigation and potential exploitation of MUM 178J to harvest useful bioactive compounds.

Keywords: *Streptomyces* sp. MUM178J; genome; anti-*Vibrio*; melanin; mangrove; SDG 3 Good health and well-being

1. Introduction

Streptomyces sp. are a group of filamentous, Gram-positive bacteria that belong to the phylum Actinobacteria ^[1, 2]. Members of the genus Streptomyces remain captivating subjects in the realm of microbiology and drug discovery as they can produce a plethora of bioactive secondary metabolites ^[3, 4]. These metabolites are produced to prevent the growth of competing microorganisms when essential nutrients are depleted ^[5-7]. This is part of the developmental process in the complex life cycle of Streptomyces sp., which ensures the survival of species ^[8, 9]. Since the discovery of the first aminoglycoside, streptomycin, from Streptomyces griseus ^[10], decades of research ensued to extract bioactive compounds from streptomycetes ^[11]. Extensive research has proven that this filamentous bacteria acts as a reservoir for bioactive secondary metabolites that have the potential for numerous applications in developing drugs and therapeutics ^[12-17]. In recent years, research on Streptomyces has shifted to isolating novel species from underexplored environments such as mangroves, deserts, oceans, and even the Arctic to search for novel bioactive compounds ^[18-27]. For example, the secondary metabolites from *Streptomyces* sp. derived from these unchartered territories have exhibited interesting bioactivities such as anticancer, antifungal, antimicrobial, cytotoxic, and antioxidant properties ^[21, 23-25, 28-34]. These findings underscore the substantial potential of the undiscovered Streptomyces species in these unique environments.

In an effort to explore the diversity of *Streptomyces* species in dynamic environments, our previous study isolated Streptomyces sp. MUM 178J from soil sampled from mangrove forests in East Malaysia ^[35]. Upon further investigation, it was found that the fermentative extract of MUM 178J inhibited the growth of a multidrug-resistant (MDR) Vibrio parahaemolyticus strain RP0132. The minimum inhibitory and minimum bactericidal concentrations were 12.5mg/mL and 50mg/mL, respectively. This interesting finding shows the potential of MUM 178J to be harnessed as an anti-Vibrio agent to control MDR V. parahaemolyticus in aquatic environments. MDR V. parahaemolyticus remains prevalent in our surrounding environment, evidenced by various studies worldwide reporting on its prevalence in clinical, environmental, and seafood samples ^[36-47]. This pathogen can be easily transmitted to humans by consuming contaminated seafood and water, causing gastroenteritis or wound infections ^[48-51]. The high prevalence of MDR V. parahaemolyticus in the environment is concerning as these bacteria will propagate and further spread antibiotic resistance (AMR) intra- and interspecies ^[52-58]. This jeopardizes the sanctity of public health as infections caused by MDR bacteria are more difficult to treat and could ultimately result in longer durations of hospitalization and higher mortality rates ^[59, 60]. As reports of MDR V. parahaemolyticus continue to surge, it is imperative to develop alternatives to antibiotics to manage this pressing issue ^[61]. Hence, discovering anti-Vibrio properties in MUM 178J provides an optimistic outlook for developing an anti-Vibrio agent that can manage MDR V. parahaemolyticus populations. This, in turn, will reduce the spread of AMR in the environment, thereby preserving the efficacy of the antimicrobial agents that are currently available for use. To better understand the underlying mechanisms of the anti-Vibrio properties of Streptomyces sp. MUM 178J, the isolate was subjected to complete whole genome sequencing. This aims to identify the biosynthetic gene clusters (BGC) responsible for the production of the bioactive compound(s) associated with its anti-Vibrio activity through genome mining and to gain insight into the genomic characteristics of MUM 178J [62-66].

2. Data description

MUM 178J was grown routinely on International *Streptomyces* Project (ISP) 2 medium at 28°C. Genomic DNA of MUM 178J was extracted using MasterPure Gram Positive DNA Purification Kit (LGC Biosearch Technologies) according to the manufacturer's instructions ^[56, 67]. The DNA quality and quantity were checked using agarose gel electrophoresis and Qubit 2.0 Fluorometer (Life Technologies, Carlsbad, CA, USA). SMRTbell DNA libraries were generated according to standard protocols and checked with Qubit for quantification and bioanalyzer for size distribution detection. The whole genome of MUM 178J was sequenced on PacBio Sequel II/IIe systems. Upon sequencing, the raw reads were assembled using Falcon, which is based on the hierarchical genome assembly and annotation completeness with single-copy orthologs. Assembled genome was annotated by NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v6.6. Ribosome RNA (rRNA) genes were analyzed by the RNAmmer and transfer RNA (tRNA) genes were predicted by the tRNAscan-SE. The whole genome of MUM 178J consists of 1 contig with G+C content

of 71.3%, the genome size of 6,699,249 bp, and a genome coverage of 300-fold. A total of 5,952 predicted genes and 5,693 protein-coding genes were detected in the genome. Based on the predictions, 69 tRNA genes, 18 rRNA genes, and 171 pseudogenes were present in the whole genome of MUM 178J (Table 1).

Streptomyces sp. MUM 178J		
Genome size (bp)	6,699,249	
Total number of contigs	1	
Contigs N ₅₀ (bp)	6,699,249	
G+C content (%)	71.3	
Genome coverage	300x	
Number of chromosomes	1	
Total number of predicted genes	5,952	
Total number of protein coding genes	5,693	
Total number of tRNA-coding genes	68	
Total number of rRNA-coding genes	18	
Total number of pseudogenes	171	

Table 1. Genomic features of *Streptomyces* sp. MUM 178J.

Furthermore, analysis based on antibiotics & Secondary Metabolite Analysis SHell (antiSMASH version 7.0) database was performed on the whole genome to detect the presence of BGC in MUM 178J^[68, 69]. This aimed to detect the presence of BGCs related to the production of secondary metabolites MUM 178J, which elicits anti-Vibrio properties. From the antiSMASH analysis, the BGC associated with the production of melanin was detected within the genome of MUM 178. Interestingly, studies have shown that melanin produced from Streptomyces sp. can exhibit anti-Vibrio properties. For instance, the melanin compound dihydroxyphenylalanine (DOPA), is produced from Streptomyces sp. MVSC6, isolated from marine sediments, elicited antagonistic activity against V. parahaemolyticus ^[70]. Moreover, marine *Streptomyces* have been reported to produce melanin pigments, which have antibacterial effects against Gram-negative bacteria, including members of the Vibrio family, such as *Vibrio cholerae*^[71]. In addition, melanin pigments produced by marinederived *Streptomyces* sp. MVCS13 was reported to inhibit fish pathogens such as *Vibrios*^[72]. Therefore, the anti-Vibrio properties of MUM 178J could be attributed to the presence of melanin BGCs in its whole genome. These findings indicate the potential of MUM 178J as an anti-Vibrio agent that can be useful in managing MDR V. parahaemolyticus in the environment in efforts to reduce AMR. The availability of the whole genome sequence of MUM 178J also allows for further studies on the strain, including genomic manipulation to produce beneficial bioactive compounds.

The whole genome sequence of MUM 178J has been deposited at DDBJ/EMBL/GenBank under accession number CP140097. The data are publicly available

at NCBI GenBank under the BioProject accession number PRJNA679911, and the BioSample accession number SAMN16862319.

Author Contributions: K-YL conducted the laboratory research work, data analysis and manuscript writing. K-OC, JW-FL and L-HL provided support on the data analysis and data management. LT-HT, JW-FL, PP, K-GC, L-HL and VL provided resources, supervision, proofreading and technical support. VL, JW-FL and L-HL conceptualized and founded this writing project. All authors have read and agreed to the published version of the manuscript.

Funding: This work is supported by the Jeffrey Cheah School of Medicine and Health Sciences Strategic Grant 2021 (Vote Number: STG-000051) awarded to VL and L-HL.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Anandan R, Dharumadurai D, and Manogaran GP. *An introduction to actinobacteria*, in *Actinobacteria-basics and biotechnological applications*. 2016, IntechOpen.
- 2. Donald L, Pipite A, Subramani R, *et al.* Streptomyces: Still the biggest producer of new natural secondary metabolites, a current perspective. Microbiol Res 2022; 13(3): 418-465.
- 3. Harir M, Bendif H, Bellahcene M, et al. Streptomyces secondary metabolites, in Basic biology and applications of actinobacteria. 2018. p. 99-122.
- 4. Challis GL and Hopwood DA. Synergy and contingency as driving forces for the evolution of multiple secondary metabolite production by Streptomyces species. PNAS 2003; 100(suppl_2): 14555-14561.
- 5. van Wezel GP and McDowall KJ. The regulation of the secondary metabolism of Streptomyces: new links and experimental advances. Nat Prod Rep 2011; 28(7): 1311-1333.
- 6. Shepherdson EMF, Baglio CR, and Elliot MA. Streptomyces behavior and competition in the natural environment. Curr Opin Microbiol 2023; 71: 102257.
- 7. Lee N, Kim W, Chung J, *et al.* Iron competition triggers antibiotic biosynthesis in Streptomyces coelicolor during coculture with Myxococcus xanthus. ISME J 2020; 14(5): 1111-1124.
- 8. Čihák M, Kameník Z, Šmídová K, *et al.* Secondary Metabolites Produced during the Germination of Streptomyces coelicolor. Front Microbiol 2017; 8.
- 9. Beroigui O and Errachidi F. Streptomyces at the heart of several sectors to support practical and sustainable applications: A review. Prog Microbes Mol Biol 2023; 6(1).
- Waksman SA, Reilly HC, and Johnstone DB. Isolation of streptomycin-producing strains of Streptomyces griseus. J Bacteriol 1946; 52(3): 393-397.
- 11. Law JW-F, Pusparajah P, Ab Mutalib N-S, *et al.* A review on mangrove actinobacterial diversity: the roles of Streptomyces and novel species discovery. Prog Microbes Mol Biol 2019; 2(1).
- 12. Antoraz S, Santamaría RI, Díaz M, *et al.* Toward a new focus in antibiotic and drug discovery from the Streptomyces arsenal. Front Microbiol 2015; 6: 461.
- 13. Barbuto Ferraiuolo S, Cammarota M, Schiraldi C, *et al.* Streptomycetes as platform for biotechnological production processes of drugs. Appl Microbiol Biotechnol 2021; 105: 551-568.
- 14. Liu R, Deng Z, and Liu T. Streptomyces species: Ideal chassis for natural product discovery and overproduction. Metab Eng 2018; 50: 74-84.

- Kemung HM, Tan LT-H, Chan K-G, *et al.* Streptomyces sp. strain MUSC 5 from mangrove forest in Malaysia: Identification, antioxidant potential and chemical profiling of its methanolic extract. Prog Microbes Mol Biol 2020; 3(1).
- 16. Tan LT-H, Lee L-H, and Goh B-H. Critical review of fermentation and extraction of anti-Vibrio compounds from Streptomyces. Prog Microbes Mol Biol 2020; 3(1).
- 17. Thye AY-K, Letchumanan V, Tan LT-H, *et al.* Malaysia's Breakthrough in Modern Actinobacteria (MOD-ACTINO) Drug Discovery Research. Prog Microbes Mol Biol 2022; 5(1).
- 18. Law JW-F, Letchumanan V, Tan LT-H, *et al.* The rising of "modern actinobacteria" era. Prog Microbes Mol Biol 2020; 3(1).
- 19. Biswas K, Choudhury JD, Mahansaria R, *et al.* Streptomyces euryhalinus sp. nov., a new actinomycete isolated from a mangrove forest. J Antibiot 2017; 70(6): 747-753.
- 20. Satheeja SV and Jebakumar SR. Phylogenetic analysis and antimicrobial activities of Streptomyces isolates from mangrove sediment. J Basic Microbiol 2011; 51(1): 71-79.
- 21. Dharmaraj S. Marine Streptomyces as a novel source of bioactive substances. World J Microbiol Biotechnol 2010; 26: 2123-2139.
- 22. Risdian C, Landwehr W, Rohde M, *et al.* Streptomyces bathyalis sp. nov., an actinobacterium isolated from the sponge in a deep sea. Antonie Van Leeuwenhoek 2021; 114: 425-435.
- 23. Sivakala KK, Gutiérrez-García K, Jose PA, *et al.* Desert environments facilitate unique evolution of biosynthetic potential in Streptomyces. Molecules 2021; 26(3): 588.
- 24. Hamid ME, Mahgoub A, Babiker AJ, *et al.* Isolation and identification of Streptomyces spp. from desert and savanna soils in Sudan. Int J Environ Res Public Health 2020; 17(23): 8749.
- 25. Liao L, Chen R, Jiang M, *et al.* Bioprospecting potential of halogenases from Arctic marine actinomycetes. BMC Microbiol 2016; 16(1): 1-9.
- 26. Sarmiento-Vizcaíno A, Martín J, Ortiz-López FJ, *et al.* Natural products, including a new caboxamycin, from Streptomyces and other Actinobacteria isolated in Spain from storm clouds transported by Northern winds of Arctic origin. Front Chem 2022; 10: 948795.
- 27. Ab Mutalib N-S, Wong SH, Ser H-L, *et al.* Bioprospecting of microbes for valuable compounds to mankind. Prog Microbes Mol Biol 2020; 3(1).
- 28. Law JW-F, Law LN-S, Letchumanan V, *et al*. Anticancer drug discovery from microbial sources: The unique mangrove streptomycetes. Molecules 2020; 25(22): 5365.
- 29. Yuan G, Hong K, Lin H, *et al.* New azalomycin F analogs from mangrove Streptomyces sp. 211726 with activity against microbes and cancer cells. Mar Drugs 2013; 11(3): 817-829.
- Law JW-F, Chan K-G, He Y-W, *et al.* Diversity of Streptomyces spp. from mangrove forest of Sarawak (Malaysia) and screening of their antioxidant and cytotoxic activities. Sci Rep 2019; 9(1): 15262.
- 31. Law JW-F, Ser H-L, Ab Mutalib N-S, *et al.* Streptomyces monashensis sp. nov., a novel mangrove soil actinobacterium from East Malaysia with antioxidative potential. Sci Rep 2019; 9(1): 3056.
- 32. Chen C, Ye Y, Wang R, *et al.* Streptomyces nigra sp. nov. is a novel actinobacterium isolated from mangrove soil and exerts a potent antitumor activity in vitro. Front Microbiol 2018; 9: 1587.
- Elabbasy EG, Hussain AA, Ashour SM, *et al.* Antifungal activity of Streptomyces canescens MH7 isolated from mangrove sediment against some dermatophytes. J Sci Res 2021; 38(part 2 (Biological Sciences)): 36-59.

- 34. Elsalam RM, Goh KW, Mahadi M, *et al.* The Antibacterial Activities of Secondary Metabolites Derived from Streptomyces sp. Prog Microbes Mol Biol 2022; 5(1).
- 35. Loo K-Y, Tan LT-H, Law JW-F, *et al.* Detection of multidrug resistant Vibrio parahaemolyticus and anti-Vibrio Streptomyces sp. MUM 178J. Prog Microbes Mol Biol 2023; 6(1).
- 36. Letchumanan V, Yin W-F, Lee L-H, *et al.* Prevalence and antimicrobial susceptibility of Vibrio parahaemolyticus isolated from retail shrimps in Malaysia. Front Microbiol 2015; 6: 33.
- 37. Letchumanan V, Pusparajah P, Tan LT-H, *et al.* Occurrence and antibiotic resistance of Vibrio parahaemolyticus from shellfish in Selangor, Malaysia. Front Microbiol 2015; 6: 1417.
- Venggadasamy V, Tan LT-H, Law JW-F, *et al.* Incidence, antibiotic susceptibility and characterization of Vibrio parahaemolyticus isolated from seafood in selangor, Malaysia. Prog Microbes Mol Biol 2021; 4(1).
- 39. Li Y, Xie T, Pang R, *et al.* Food-borne Vibrio parahaemolyticus in China: prevalence, antibiotic susceptibility, and genetic characterization. Front Microbiol 2020; 11: 1670.
- 40. Li H, Tang R, Lou Y, *et al.* A comprehensive epidemiological research for clinical Vibrio parahaemolyticus in Shanghai. Front Microbiol 2017; 8: 1043.
- 41. Ceccarelli D, Hasan NA, Huq A, *et al.* Distribution and dynamics of epidemic and pandemic Vibrio parahaemolyticus virulence factors. Front Cell Infect 2013; 3: 97.
- 42. Igbinosa EO, Beshiru A, Igbinosa IH, *et al.* Prevalence and characterization of food-borne Vibrio parahaemolyticus from African salad in southern Nigeria. Front Microbiol 2021; 12: 632266.
- 43. Xie T, Xu X, Wu Q, *et al.* Prevalence, molecular characterization, and antibiotic susceptibility of Vibrio parahaemolyticus from ready-to-eat foods in China. Front Microbiol 2016; 7: 549.
- 44. Esteves K, Mosser T, Aujoulat F, *et al.* Highly diverse recombining populations of Vibrio cholerae and Vibrio parahaemolyticus in French Mediterranean coastal lagoons. Front Microbiol 2015; 6.
- 45. Chen AJ, Hasan NA, Haley BJ, *et al.* Characterization of Pathogenic Vibrio parahaemolyticus from the Chesapeake Bay, Maryland. Front Microbiol 2017; 8.
- 46. Chen L, Wang J, Chen J, *et al.* Epidemiological characteristics of Vibrio parahaemolyticus outbreaks, Zhejiang, China, 2010–2022. Front Microbiol 2023; 14.
- 47. Narayanan SV, Joseph TC, Peeralil S, *et al.* Prevalence, Virulence Characterization, AMR Pattern and Genetic Relatedness of Vibrio parahaemolyticus Isolates From Retail Seafood of Kerala, India. Front Microbiol 2020; 11.
- 48. Letchumanan V, Chan K-G, and Lee L-H. Vibrio parahaemolyticus: a review on the pathogenesis, prevalence, and advance molecular identification techniques. Front Microbiol 2014; 5: 705.
- 49. Loo K-Y, Law JW-F, Tan LT-H, *et al.* The burden of Vibrio sp. infections–A scoping review. Prog Microbes Mol Biol 2023; 6(1).
- 50. Raghunath P. Roles of thermostable direct hemolysin (TDH) and TDH-related hemolysin (TRH) in Vibrio parahaemolyticus. Front Microbiol 2015; 5: 805.
- 51. Tan CW, Malcolm TTH, Kuan CH, *et al.* Prevalence and Antimicrobial Susceptibility of Vibrio parahaemolyticus Isolated from Short Mackerels (Rastrelliger brachysoma) in Malaysia. Front Microbiol 2017; 8.
- 52. Lee L-H, Ab Mutalib N-S, Law JW-F, *et al.* Discovery on antibiotic resistance patterns of Vibrio parahaemolyticus in Selangor reveals carbapenemase producing Vibrio parahaemolyticus in marine and freshwater fish. Front Microbiol 2018; 9: 2513.

- 53. Letchumanan V, Ser H-L, Tan W-S, *et al.* Genome sequence of Vibrio parahaemolyticus VP152 strain isolated from Penaeus indicus in Malaysia. Front Microbiol 2016; 7: 1410.
- 54. Letchumanan V, Ser H-L, Chan K-G, *et al.* Genome sequence of Vibrio parahaemolyticus VP103 strain isolated from shrimp in Malaysia. Front Microbiol 2016; 7: 1496.
- 55. Letchumanan V, Tan W-S, Yin W-F, *et al.* Genome sequence of Vibrio sp. OULL4 isolated from shellfish. Prog Microbes Mol Biol 2020; 3(1).
- 56. Loo K-Y, Tan LT-H, Law JW-F, *et al.* Complete Whole Genome Sequence of Vibrio parahaemolyticus RP0132 Strain Isolated from Shrimp in Malaysia. Prog Microbes Mol Biol 2023; 6(1).
- 57. da Silva LV, Ossai S, Chigbu P, *et al.* Antimicrobial and Genetic Profiles of Vibrio vulnificus and Vibrio parahaemolyticus Isolated From the Maryland Coastal Bays, United States. Front Microbiol 2021; 12.
- 58. Dutta D, Kaushik A, Kumar D, *et al.* Foodborne Pathogenic Vibrios: Antimicrobial Resistance. Front Microbiol 2021; 12.
- Tang KWK, Millar BC, and Moore JE. Antimicrobial resistance (AMR). Br J Biomed Sci 2023; 80: 11387.
- 60. de Kraker MEA and Lipsitch M. Burden of Antimicrobial Resistance: Compared to What? Epidemiol Rev 2021; 43(1): 53-64.
- 61. Goh JXH, Tan LT-H, Law JW-F, *et al.* Probiotics: Comprehensive Exploration of the Growth Promotion Mechanisms in Shrimps. Prog Microbes Mol Biol 2023; 6(1).
- 62. Ser H-L, Law JW-F, Tan W-S, *et al.* Whole genome sequence of Streptomyces colonosanans strain MUSC 93JT isolated from mangrove forest in Malaysia. Prog Microbes Mol Biol 2020; 3(1).
- 63. Ser H-L, Tan LT-H, Tan W-S, *et al.* Whole-genome sequence of bioactive streptomycete derived from mangrove forest in Malaysia, Streptomyces sp. MUSC 14. Prog Microbes Mol Biol 2021; 4(1).
- 64. Law JW-F, Letchumanan V, Hong K-W, *et al.* Streptomyces learnhanii sp. nov., unveiling a Mangrove-Derived Novel "Modern Actinobacteria" in Malaysia. Prog Microbes Mol Biol 2023; 6(1).
- 65. Law JWF, Tan LT-H, Letchumanan V, *et al.* Streptomyces griseiviridis sp. nov., a Novel "Modern Actinobacteria" isolated from Malaysia Mangrove Soil. Prog Microbes Mol Biol 2023; 6(1).
- 66. Pusparajah P, Law JW-F, Chan K-G, *et al.* Whole-Genome Sequence of Streptomyces pluripotens strain MUM 16J, a Potential Resource of Glycopeptide Antibiotic and Biocontrol Agent against Biofilm-forming Bacteria. Prog Microbes Mol Biol 2023; 6(1).
- 67. Lino FSdO, Misiakou M-A, Kang K, *et al.* Strain dynamics of specific contaminant bacteria modulate the performance of ethanol biorefineries. bioRxiv 2021: 2021.02. 07.430133.
- 68. Blin K, Medema MH, Kottmann R, *et al.* The antiSMASH database, a comprehensive database of microbial secondary metabolite biosynthetic gene clusters. Nucleic Acids Res 2016: gkw960.
- 69. Weber T, Blin K, Duddela S, *et al.* antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res 2015; 43(W1): W237-W243.
- Sivaperumal P, Kamala K, and Rajaram R. Bioactive DOPA melanin isolated and characterised from a marine actinobacterium Streptomyces sp. MVCS6 from Versova coast. Nat Prod Res 2015; 29(22): 2117-2121.
- 71. Vasanthabharathi V, Lakshminarayanan R, and Jayalakshmi S. Melanin production from marine Streptomyces. Afr J Biotechnol 2011; 10(54): 11224.

72. Sivaperumal P, Kamala K, Rajaram R, *et al.* Melanin from marine Streptomyces sp. (MVCS13) with potential effect against ornamental fish pathogens of Carassius auratus (Linnaeus, 1758). Biocatal Agric Biotechnol 2014; 3(4): 134-141.



Author(s) shall retain the copyright of their work and grant the Journal/Publisher right for the first publication with the work simultaneously licensed under:

Creative Commons Attribution-NonCommercial 4.0 International (CC BY-NC 4.0). This license allows for the copying, distribution and transmission of the work, provided the correct attribution of the original creator is stated. Adaptation and remixing are also permitted.