

A Health Impact Assessment of CRISPR-Cas9 as a Therapeutic Approach for Prader-Willi Syndrome

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Abstract: Prader-Willi Syndrome is a rare neurodevelopmental genetic disorder resulting from expression defects of the exclusively paternally-expressed PWS locus at chromosome 15q11-q13. Even though its multifaceted manifestations and potentially fatal comorbidities poses universal concern, it remains incurable. Thus, this commentary applies a health impact assessment framework to evaluate the potential behind CRISPR-Cas9 gene-editing technology as a therapeutic intervention for PWS. While this novel technique displays outstanding precision and effectiveness in the targeted identification and demethylation of the silenced maternal locus, challenges such as off-target mutations, safety, and ethical implications warrant additional improvements. Therefore, recommendations regarding future research and clinical implementation are also proposed.

Keywords: CRISPR-Cas9; Gene-editing; Health impact assessment; Prader-Willi Syndrome

1. Introduction

Prader-Willi Syndrome (PWS) is a condition of rare genetic origin due to an imprinting defect on chromosome 15q11-q13 that results in a variety of developmental, metabolic, neurological, and behavioral abnormalities^[1]. The condition is most commonly associated with loss of the paternal PWS Imprinting Centre (PWS-IC), which comprises the small nucleolar RNA SNORD116 gene cluster within the SNRPN transcript^[2]. In addition to a high risk for comorbidities like obesity, clinical presentations vary from intellectual disabilities, growth anomalies, and feeding disorders, based on the severity of chromosomal defects^[3].

In addition, the prevalence of PWS occurs in 1 out of each 20,000 live births, and fatality rate each year is 3%^[1]. The median age of diagnosis is still at 0.3 years, although there exist some cases occurring later in life, though earlier diagnosis is highly important to support effective disease regulation^[2]. Furthermore, since no cure for PWS has been developed so far, new therapeutic strategies to the PWS genetic defect must be investigated, particularly in light of the fact that current treatments are mostly symptomatic. Clustered Regularly Interspaced Short Palindromic Repeats-CRISPR associated protein 9 (CRISPR-Cas9) gene editing is a potential treatment based on the high disease burden. This is due to the fact that it reactivates the epigenetically silenced maternal allele, including its PWS locus, in a specific way^[4]. A health impact assessment (HIA) framework is therefore employed in this commentary to explore the feasibility of CRISPR-Cas9 in PWS management.

2. Health Impact Assessment: Evaluating the Potential of CRISPR-Cas9

A HIA provides a structured and systematic method in predicting and evaluating the health implications of a proposed intervention, policy, or technology^[5,6]. Within the context of CRISPR-Cas9 as a potential approach for PWS, a HIA allows for a comprehensive assessment of anticipated benefits, risks, and ethical concerns associated with this gene-editing technology. Considering the novelty of CRISPR-Cas9 in PWS management, an observational epidemiological study design using existing scientific literature proves as a suitable evaluation framework.

The HIA process involves key steps: screening, scoping, risk assessment, and recommendations. Screening involves a preliminary assessment behind the necessity of a HIA, which is evident given the substantial health burden of PWS and the promising recent advancements of CRISPR-Cas9 as a breakthrough intervention^[1,3]. Meanwhile, scoping identifies critical areas of evaluation, like measurable health outcomes including diagnostic timelines and hospitalisation rates. A baseline profile highlights poor conditions with early stage diagnostic challenges^[2,7], and deteriorating physical and psychological health status of PWS patients as studies cite overlooked PWS-related comorbidities in adulthood and increasing suicidality^[8,9]. Comparatively, reports of CRISPR-Cas9's ability to accurately pinpoint and treat the root cause of PWS within the genome provide strong indication and promise for reducing negative health impacts and improving patient outlook. Table 1 presents a consolidated overview of the HIA conducted for the application of CRISPR-Cas9 in Prader-Willi Syndrome, integrating the key stages of the HIA process with associated clinical benefits, potential risks, ethical considerations, and recommended future actions.

Table 1. Summary of the Health Impact Assessment (HIA) of CRISPR-Cas9 Gene-Editing for Prader-Willi Syndrome

HIA Step	Focus Area	Key Points
Screening	Necessity of Evaluation	High mortality (3% annually); significant developmental, behavioural, and metabolic burden; lack of curative therapies.
Scoping	Health Outcomes	Delayed diagnosis (median age 0.3 yrs), overlooked adult comorbidities, increasing mental health risks (e.g., suicidality).
Risk Assessment	Potential Benefits	- Precise reactivation of maternal SNORD116. - Compensation for absent paternal locus. - Avoids UBE3A disruption (prevents Angelman Syndrome).
	Risks & Challenges	- Off-target effects on MKRN3, MAGEL2, NDN genes. - Risk of inducing syndromes like Schaaf-Yang. - Neurodevelopmental disruptions possible.
	Ethical Principles	- Autonomy: Informed consent essential. - Transparency: Risks/benefits must be clearly communicated. - Equity: Ensure access across populations. - Regulation: Varying global laws; need for oversight.
Recommendations	Strategic Actions	- Enhance CRISPR targeting and delivery systems. - Combine with symptom-based therapies. - Conduct clinical trials. - Establish ethical guidelines.

3. Potential Benefits, Risks and Ethical Considerations of CRISPR-Cas9 in PWS

3.1 Potential Benefits

CRISPR-Cas9 presents an efficient approach for overcoming the imprinting defect underlying PWS. Following the highly specific identification, targeted DNA demethylation and transient reactivation of the intact but epigenetically silenced maternal allele, the brief maternal PWS-IC expression, containing the SNORD116 gene cluster in the SNRPN transcript, is sufficient to compensate for the absent paternal locus, as demonstrated in in vitro models^[4]. Moreover, the short term reactivation of the maternal PWS locus showed no interference to the maternally expressed UBE3A antisense transcript distal to the SNRPN transcript in iPSC-derived neuronal cells even through neuronal differentiation, thereby avoiding the risk of the emergence of Angelman Syndrome^[4,10].

3.2 Risks and Challenges

Despite its potential, the implementation of CRISPR-Cas9 for PWS poses significant uncertainties. The presence of other paternally expressed genes (e.g., MKRN3, MAGEL2, NDN) near the SNRPN/SNORD116 cluster increases the risk of off-target editing as shown in Figure 1. Unintentional reactivation of these regions may aggravate phenotypic complications. One major concern is the possibility of off-target effects, particularly on adjacent highly regulated regions within the PWS-IC, such as the MKRN3, MAGEL2, and NDN genes that share the monoallelic paternal expression of the SNRPN transcript^[11,12]. Non-expression of these genes jeopardise major neurological processes and contribute to characteristic PWS symptoms, namely central precocious puberty with accompanying obesity and behavioural aberrations, Schaaf-Yang Syndrome affecting development of the central nervous system, and sleep-wake disturbances leading to developmental delays, mental deficiencies and behaviour problems^[2]. Yet since these genes are not the primary cause for PWS, reactivation may produce unnecessary risks for the patient, or may even cause more harm than good. Thus, a cautious approach is required to ensure that gene editing is restricted to the SNRPN host transcript, minimizing unintended consequences.

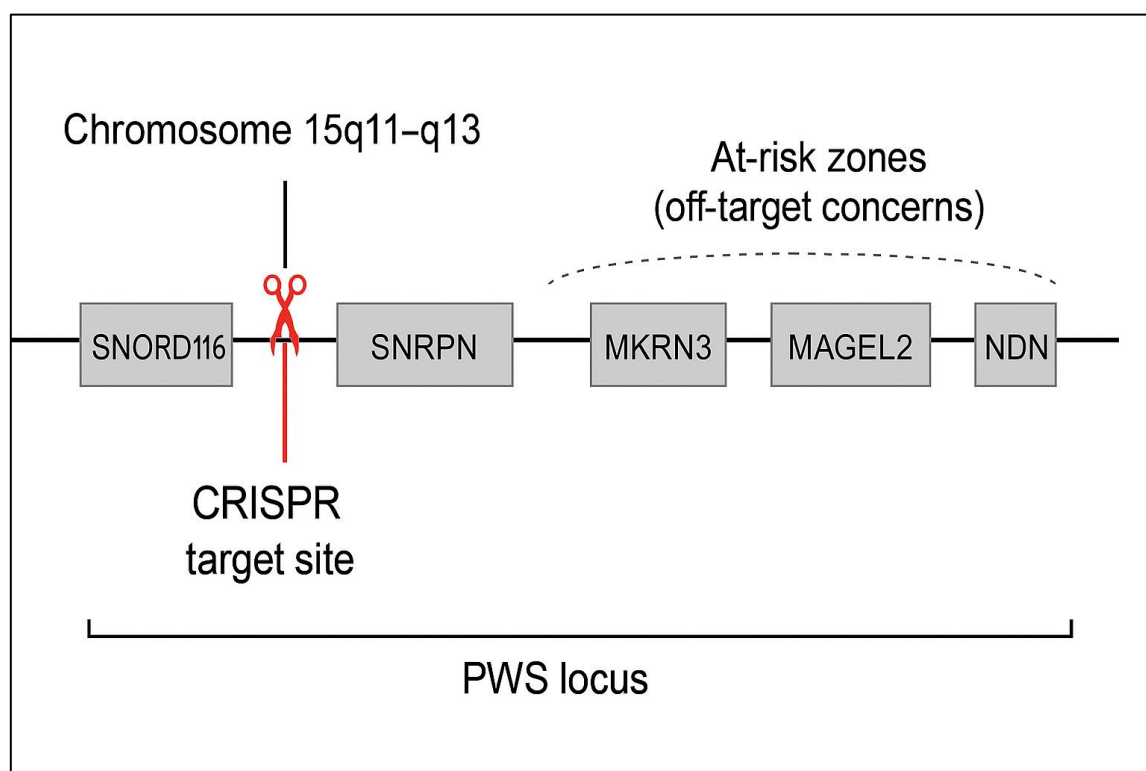


Figure 1. Schematic of the PWS-critical region on chromosome 15q11-q13. The CRISPR-Cas9 target site (SNRPN/SNORD116) is highlighted, with adjacent genes (MKRN3, MAGEL2, NDN) flagged as potential off-target concerns due to their paternal imprinting and neurological roles.

3.3 Ethical Considerations

Applying CRISPR-Cas9 technology to human genetic disorders requires careful consideration of ethical issues^[13]. Upholding a number of ethical standards is necessary, including:

- Respect for autonomy: Individuals or their legal guardians are to provide informed consent before gene-editing procedures are carried out.
- Transparency: Open disclosure of risks, benefits, and uncertainties associated with CRISPR-Cas9 for PWS.
- Equitable access: As PWS is a condition that occurs in people of all ages, sexes, and races, ensuring fair access to gene-editing treatments is essential to preventing health inequalities.
- Regulatory oversight: Countries possess different levels of regulatory power and acceptability towards novel gene-editing technologies, and there are ongoing controversies between somatic and germline engineering.

Resolution of these ethical challenges demands a strong framework with an emphasis on protecting patients but also encouraging responsible innovation.

4. Recommendations for Future Research and Clinical Translation

The following recommendations must be taken into consideration to increase the therapeutic potential of CRISPR-Cas9 for PWS and to reduce the risks involved:

- Enhanced targeting: Further CRISPR-based epigenetic screens are recommended to improve the specificity and efficacy of SNRPN host transcript targeting^[4].
- Optimized delivery mechanisms: For in vivo applications, delivery vectors that are compatible with the large molecular weight of dCas9-based epigenetic editors are crucial to achieve maximum efficacy of treatment as well as fewer side effects^[4].
- Combination therapies: Coupling CRISPR-Cas9 with current therapeutic interventions, such as hormone replacement therapy or drug supplementation, to treat the symptoms associated with MKRN3, MAGEL2, and NDN can be a more effective and less invasive treatment approach^[14,15].
- Clinical trials: In order to refine precision methods and identify the long-term effects, safety, and effectiveness of CRISPR-Cas9 therapies in PWS patients, well-controlled clinical trials are required.
- Robust ethical guidelines: Formulating strict ethical frameworks for the governance of clinical gene editing applications is central to ensuring adherence to the ethical principles of informed consent, transparency, and equitable access^[13].

5. Conclusion

CRISPR-Cas9 is a revolutionary technique of therapeutic intervention in the genetic etiology of PWS. Preclinical models show promising results; yet, before this technology is used in medicine, long-term safety, efficacy, and ethical issues need to be widely examined. Its safe and efficient use can be ensured by improved vector systems, multimodal therapy, additional clinical trials, enhanced gene targeting and epigenetic screening, and rigid adherence to ethical guidelines. Researchers and medical professionals can help establish a promising and innovative genetic treatment for patients with PWS by prioritizing these steps, which will ultimately enhance their quality of life and future health prospects.

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References

1. Kummerfeld DM, Raabe CA, Brosius J, *et al.* A Comprehensive Review of Genetically Engineered Mouse Models for Prader-Willi Syndrome Research. *IJMS* 2021; 22(7): 3613.
2. Szabadi S, Sila Z, Dewey J, *et al.* A Review of Prader–Willi Syndrome. *Endocrines* 2022; 3(2): 329–348.
3. Butler MG, Oyetunji A, Manzardo AM. Age Distribution, Comorbidities and Risk Factors for Thrombosis in Prader–Willi Syndrome. *Genes* 2020; 11(1): 67.
4. Rohm D, Black JB, McCutcheon SR, *et al.* Activation of the imprinted Prader-Willi Syndrome locus by CRISPR-based epigenome editing [Internet]. 2024 [cited 2024 May 31]; Available from: <http://biorxiv.org/lookup/doi/10.1101/2024.03.03.583177>
5. Olalekan RM, Adindu IB, Udensi EO, *et al.* Health Impact Assessment: Expanding Public Policy Tools for Promoting Sustainable Development Goals (SDGs) in Nigeria. *Health Impact Assessment* 2020;
6. Thondoo M, Mueller N, Rojas-Rueda D, *et al.* Participatory quantitative health impact assessment of urban transport planning: A case study from Eastern Africa. *Environ Int* 2020; 144: 106027.
7. Grootjen LN, Uyl NEM, Van Beijsterveldt IALP, *et al.* Prenatal and Neonatal Characteristics of Children with Prader-Willi Syndrome. *JCM* 2022; 11(3): 679.
8. Peleggi A, Bohonowych J, Strong TV, *et al.* Suicidality in individuals with Prader-Willi syndrome: a review of registry survey data. *BMC Psychiatry* 2021; 21(1): 438.
9. Pellikaan K, Rosenberg AGW, Kattentidt-Mouravieva AA, *et al.* Missed Diagnoses and Health Problems in Adults With Prader-Willi Syndrome: Recommendations for Screening and Treatment. *J Clin Endocrinol Metab* 2020; 105(12): e4671–e4687.
10. Gilmore RB, Gorka D, Stoddard CE, *et al.* Generation of isogenic models of Angelman syndrome and Prader-Willi syndrome in CRISPR/Cas9-engineered human embryonic stem cells [Internet]. 2023 [cited 2024 May 23]; Available from: <http://biorxiv.org/lookup/doi/10.1101/2023.08.30.555563>

11. Duan Y, Liu L, Zhang X, *et al.* Phenotypic spectrum and mechanism analysis of Schaff Yang syndrome: A case report on new mutation of MAGEL2 gene. *Medicine* 2021;100(24): e26309.
12. Li J, Chen W, Li D, *et al.* Conservation of Imprinting and Methylation of MKRN3, MAGEL2 and NDN Genes in Cattle. *Animals* 2021; 11(7): 1985.
13. Gostinskaya I. CRISPR–Cas9: A History of Its Discovery and Ethical Considerations of Its Use in Genome Editing. *Biochemistry Moscow* 2022;87(8):777–88.
14. Althammer F, Muscatelli F, Grinevich V, *et al.* Oxytocin-based therapies for treatment of Prader-Willi and Schaaf-Yang syndromes: evidence, disappointments, and future research strategies. *Transl Psychiatry* 2022; 12(1): 318.
15. Bohonowych J, Miller J, McCandless SE, *et al.* The Global Prader–Willi Syndrome Registry: Development, Launch, and Early Demographics. *Genes* 2019; 10(9): 713.



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